

Gel Analysis of acidic proteins by Stains All

Take 30 ul of sample (media or cells scraped into PBS and sonicated).

Add 10 ul of sample buffer

Use 5 ul rainbow markers

Run in an acrylamide gel (10%, 12%, 4-20, 4-12% SDS/MOPs from novex) (take white tape off!). Fill gel chamber more than half way outside and all the way to the top inside.

**Run 125 volts/8 watts until tracking dye approaches the bottom of the gel
Crack plastic to release the gel. Ease it into isopropanol (use heavy spatula and old X-ray film for support)**

Wash with isopropanol 4 times with overnight incubation or alternate isopropanol with water at least 4 times during a two hour period

Stain 1 hour in "stains All" (shield from the light)

Rinse and take a picture

Dry gel 50 C for 3 hours (if stained in code blue/coomassie gel type stain, use 80 C for 1 hour)

Stains All:

30 ml stock

30 formamide

150ml isopropanol

6 ml 1.5 M tris Ph8.8

to 600 ml with water

Stock: 0.1% Stains All in fresh formamide